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Synthesis of optically active oxazoles from phosphorylated 2*H*-azirines and *N*-protected amino acids or peptides

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Abstract—A simple synthesis of optically active phosphorylated oxazoles 8, 10, 11, and 12 containing amino acid residues from 2*H*-azirine-phosphine oxides 1 or -phosphonates 6 is described. Ring-opening reaction of 2*H*-azirines derived from phosphine oxides 1 and phosphonates 6 with *N*-protected amino acids 2 gives functionalized phosphorylated ketamides 3 and 7. Cyclization of ketamides 3 and 7 with triphenylphosphine and hexachloroethane in the presence of triethylamine leads to the formation of racemic and optically active phosphorylated oxazoles containing *N*-protected amino acid residues 8 and 10. Deprotection of these oxazoles gives aminoalkyl oxazoles 11 and 12. 2*H*-Azirines 1 and 6 also react with *N*-protected peptides 13 and give functionalized ketamides 14 and 15, ring closure of which leads to the formation of phosphorylated oxazoles containing peptide residues 16 and 17. \bigcirc 2002 Elsevier Science Ltd. All rights reserved.

1. Introduction

Oxazoles are widely used intermediates for functional transformations.¹ Simple oxazoles are common units in a wide variety of polyoxazole marine natural products possessing biological activity,2 while aminoalkyl oxazoles derived from amino acids I are very important substrates because they are constituents of peptidebased alkaloids and antibiotics with remarkable cytotoxic and/or antitumor properties.^{3,4} Additionally, amino acid residues play an important role in connecting the heterocyclic moieties in the formation of polyazole natural products. Therefore, the preparation of optically active oxazoles I is an interesting goal in synthetic organic chemistry. Furthermore, it is known that phosphorus substituents regulate important biological functions,⁵ and that molecular modifications involving the introduction of organophosphorus functionalities could increase their biological activity, in a similar way to that reported for other pharmaceuticals.⁵ For these reasons, optically active oxazoles with amino acid residues, which contain a phosphorus substituent (II, Fig. 1) are expected to play a similar role to that observed in the isosteric analogues I, and could possess biological activity and be used in the preparation of new complex polyoxazole derivatives containing phosphorus substituents. However, as far as we know, no examples of racemic or optically active oxazoles with amino acid residues and containing a phosphorus substituent have been described.

In this context, we have described new methods for the preparation of five-⁶ and six-⁷ membered phosphorus substituted nitrogen heterocycles from functionalized phosphine oxides and phosphonates and the synthetic uses of amino phosphorus derivatives as starting materials for the preparation of acyclic compounds⁸ and phosphorus-containing heterocycles.⁹ Recently, we reported the first asymmetric synthesis of 2H-azirines¹⁰ derived from phosphine oxides by alkaloid-mediated Neber reaction of tosyl oximes,^{11,12} as well as their dimerization to phosphorylated pyrazines¹³ and the ring opening of azirines with carboxylic acids followed by the cyclization of the corresponding adducts to oxazoles **IV** (Scheme 1).¹⁴ Continuing with our interest in



Figure 1.

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Scheme 1.

the synthesis of new phosphorus substituted heterocycles, we report herein an easy and high yielding synthesis of racemic and optically active 4-(2-aminoalkyloxazolyl)phosphine oxides (II, Fig. 1, R = Ph) and -phosphonates (II, Fig. 1, R = OEt) from easily available azirines. Azirines are thus very interesting reagents for the activation and amidation of *N*-protected amino acids via azirine–oxazolone intermediates and have been successfully applied to peptide synthesis.^{15–17}

2. Results and discussion

2.1. Reaction of azirines 1 and 6 with *N*-protected amino acids 2. Synthesis of α -ketamides 3 and 7

Ring opening and selective cleavage of the N–C double bond of amino-azirines^{15,16} and of phosphorus substituted azirines,¹⁴ can be achieved with carboxylic acids. In the case of amino-azirines the process has been extended to *N*-protected amino acids.^{15,17} As far as we know, no ring-opening reaction of azirines containing phosphorus substituents with *N*-protected amino acid has been reported. For this reason, we explored the reaction of phosphorylated azirines with *N*-protected amino acids and peptides. Given the increasing interest in 'phosphapeptides' in organic and medicinal chemistry,^{18,19} this reaction can be used as a model for the introduction of amino phosphorus moieties into peptides. Furthermore, the functionalized ketamides generated can be used for the regioselective preparation of the previously unknown racemic and optically active phosphorylated oxazoles containing amino alkyl residues.

Firstly, we explored the reaction of azirines with racemic N-Boc protected amino acids.²⁰ Reaction of 3-methyl-2*H*-azirinyl phosphine oxide 1a $(R^1 = CH_3)$ with N-Boc-glycine 2a ($R^2 = R^3 = H$), at low temperature (-80°C) in THF led to the formation of α ketamide containing a phosphine oxide group in the α -position 3aa (R¹=CH₃, R²=R³=H) (Scheme 2, Table 1, entry 1). The reaction can be extended both to C α -substituted (±)-N-Boc-alanine **2b** (R²=CH₃, R³= H; $R^2 = H$, $R^3 = CH_2$) and to (±)-N-Boc-serine 2c ($R^2 =$ CH₂OH, R^3 =H; R^2 =H, R^3 =CH₂OH) obtaining (1:1) diastereoisomeric mixtures²¹ of α -ketamides **3ab** (R¹= CH_3 , $R^2 = CH_3$, $R^3 = H$; $R^1 = CH_3$, $R^2 = H$, $R^3 = CH_3$) and **3ac** $(R^1 = CH_3, R^2 = CH_2OH, R^3 = H; R^1 = CH_3,$ $R^2 = H, R^3 = CH_2OH$) (Scheme 2, Table 1, entries 2 and 3). Spectroscopic data were in agreement with the assigned structure of compounds 3. Mass spectrometry of **3ab** showed the molecular ion peak, while in the ${}^{31}P$ NMR spectrum phosphine oxide groups resonated at $\delta_{\rm P} = 32.4$ and 33.1 ppm for both diastereoisomers. The formation of adducts 3 could be explained by protonation of the nitrogen atom of the azirine, then nucleophilic addition of the carboxylate to the aziridinium ion, followed by ring expansion of aziridine 4 to give the zwitterionic oxazolone 5, which underwent ring opening to form ketamides 3.

The scope of the reaction was not limited to racemic N-protected amino acids **3aa–3ac**, given that azirine



Table	1.	α-Ketamides	3	and	7
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Entry	Compound	R	\mathbb{R}^1	R ²	R ³	Yield (%) ^a	$[\alpha]_{D}^{22,b}$
1	3 aa	C ₆ H ₅	CH ₃	Н	Н	73	_
2	(±)- 3ab	C_6H_5	CH ₃	Hc	CH ₃ ^c	65	_
3	(\pm) -3ac	C_6H_5	CH ₃	Hc	CH ₂ OH ^c	65	_
4	(+)- 3ad	C_6H_5	CH ₃	CH ₃	Н	58	+34.0
5	(-)- 3ae	C_6H_5	CH ₃	Н	CH ₃	62	-34.0
6	(+)- 3af	C_6H_5	CH ₃	CH ₂ OH	Н	48	+32.4
7	(-)- 3ag	C_6H_5	CH ₃	Η	CH ₂ OH	66	-32.4
8	(-)- 3ah	C_6H_5	CH ₃	Н	CH ₂ C ₆ H ₅	56	-11.4
9	(<u>+</u>)-7ab	OC_2H_5	CH ₃	Hc	CH ₃ ^c	68	_
10	(+)-7ad	OC ₂ H ₅	CH ₃	CH ₃	Н	71	+21.0
11	(-)-7ae	OC_2H_5	CH ₃	Н	CH ₃	66	-21.0
12	(-)-7ag	OC_2H_5	CH ₃	Н	CH ₂ OH	52	-11.0
13	(+)-7bd	OC_2H_5	C_2H_5	CH ₃	Н	72	+24.4
14	(–)-7bg	OC_2H_5	C_2H_5	Н	CH ₂ OH	50	-9.5

^a Yields refer to isolated compounds.

^b Degrees (for concentration, see Section 4).

^c Racemic.

phosphine oxide 1a ($R^1 = CH_3$) also reacted with optically active isomers of N-Boc-(R)-alanine 2d ($R^2 = CH_3$, $R^3 = H$) or (S)-alanine 2e ($R^2 = H$, $R^3 = CH_3$), of N-Boc-(R)-serine 2f ($R^2 = CH_2OH$, $R^3 = H$) or (S)-serine 2g ($R^2 = H$, $R^3 = CH_2OH$) and with N-Boc-(S)-phenylalanine **2h** ($R^2 = H$, $R^3 = CH_2C_6H_5$) to give, respectively, optically active α -ketamides (+)-3ad (R¹=CH₃, R²= CH_3 , $R^3 = H$), (-)-3ae ($R^1 = CH_3$, $R^2 = H$, $R^3 = CH_3$), (+)-3af ($R^1 = CH_3$, $R^2 = CH_2OH$, $R^3 = H$), (-)-3ag ($R^1 =$ CH_3 , $R^2 = H$, $R^3 = CH_2OH$) and (-)-3ah ($R^1 = CH_3$, $R^2 = H$, $R^3 = CH_2C_6H_5$), which were obtained, as in the case of the racemic compounds, as 1:1 diastereoisomeric mixtures²¹ (Scheme 2, Table 1, entries 4-8). Enantiomerically enriched azirine phosphine oxide 1a $(R^1 = CH_3)^{11}$ was also treated with optically active N-Boc-(R)- 2d or (S)-amino acids 2e and 2g but enantiomerically enriched ketamides 3 were not obtained and (1:1) diastereoisomeric mixtures²¹ were obtained instead. These results could be explained by epimerization of C α -carbon to the phosphine oxide group, due to the acid character of the methinic hydrogen in this position.

The process can also be extended to azirines derived from phosphonates 6^{12a} In this case shorter periods of time (2 h) and higher temperatures (70°C) were required. Heating racemic or enantiomerically enriched azirines **6a** ($\mathbf{R}^1 = \mathbf{C}\mathbf{H}_3$), and **6b** ($\mathbf{R}^1 = \mathbf{C}_2\mathbf{H}_5$), with N-Boc-protected- (\pm) -alanine **2b**, -(R)-alanine **2d** or -(S)alanine 2e and with N-Boc-(S)-serine 2g led to the formation²² of racemic and optically active α -ketamides containing a diethoxyphosphoryl group in the α -position 7ab-7bg, which were obtained, as before, as 1:1 diastereoisomeric mixtures²¹ (Scheme 2, Table 1, entries 9-14). This formation of phosphapeptides^{18,19} can be regarded as a peptide chain elongation which introduces an α -ketamine containing a phosphonate group or a phosphine oxide to the C-terminal end of the amino acid.

2.2. Synthesis of racemic and optically active phosphorus substituted aminoalkyl oxazoles 8 and 11

Next we explored the ring closure of ketamides containing a phosphine oxide 3, or phosphonate substituent 7 for the preparation of the previously unknown and potentially useful racemic and optically active phosphorylated oxazoles containing amino alkyl residues II (Fig. 1). The best results were observed when triphenylphosphine dichloride, generated 'in situ' from the reaction of the phosphine with hexachloroethane, was used.^{23,24} Adducts 3 were treated with triphenylphosphine and hexachloroethane in the presence of triethylamine in THF to give oxazole phosphine oxides 8 in good yields and in a regioselective fashion (Scheme 3, Table 2). Spectroscopic data were in agreement with the assigned structure of compounds 8. Mass spectrometry of (+)-8ad showed the molecular ion peak, while in the ³¹P NMR spectrum the phosphine oxide group resonated at $\delta_{\rm P} = 18.3$ ppm. The ¹³C NMR spectrum of oxazole (+)-8ad showed doublets at $\delta_{\rm C} = 126.3$ ppm (${}^{1}J_{PC}$ =141.5 Hz) for C-4, and at δ_{C} =163.8 ppm $({}^{3}J_{PC} = 17.1 \text{ Hz})$ for C-2. The formation of oxazoles 8 could be explained by deprotonation of ketamides 3 by means of dichlorotriphenylphosphorane (Ph₃PCl₂), generated in situ from triphenylphosphine and hexachloroethane,²⁵ to give an intermediate enamide 9 (Scheme 3) followed by the loss of triphenylphosphine oxide and subsequent ring closure.24

Cyclization is quite general, allowing the preparation of racemic and optically active oxazole phosphine oxides (4-position) bearing aminoalkyl groups in the 2-position of the ring derived not only from glycine **8aa** (R^1 =CH₃, R^2 = R^3 =H) (Scheme 3, Table 2, entry 1), but also from (±)-alanine **8ab** (R^1 =CH₃, R^2 =CH₃, R^3 =H; R^1 =CH₃, R^2 =H, R^3 =CH₃), (*R*)-alanine (+)-**8ad** (R^1 =CH₃, R^2 =H, R^3 =H) or (*S*)-alanine (-)-**8ae** (R^1 =CH₃, R^2 =H, R^3 =CH₃) (Scheme 3, Table 2,

entries 2–4), and from (S)-phenylalanine (–)-**8ah** ($R^1 = CH_3$, $R^2 = H$, $R^3 = CH_2C_6H_5$) (Scheme 3, Table 2, entry 5). As far as we know, this process represents the first synthesis of optically active oxazoles derived from amino acids and containing a phosphorus substituent.

Likewise the cyclization of functionalized ketamides derived from α -aminophosphonates 7 with triphenylphosphine-hexachloroethane and triethylamine gave (±)-oxazole **10ab** (R¹=CH₃, R²=CH₃, R³=H; R¹= CH₃, R²=H, R³=CH₃) (Scheme 3, Table 2, entry 6) when racemic ketamide **7ab** was used and optically active oxazole phosphonates containing aminoalkyl groups in the 2-position of the ring derived from (*R*)alanine (+)-**10ad** (R¹=CH₃, R²=CH₃, R³=H) and (+)-**10bd** (R¹=C₂H₅, R²=CH₃, R³=H) or from (*S*)-alanine (-)-**10ae** (R¹=CH₃, R²=H, R³=CH₃) (Scheme 3,

Table 2, entries 7–9) when optically active ketamides (+)-7ad, (+)-7bd, (-)-7ae were used. The deprotection of the terminal N-Boc group of oxazoles 8 and 10 was studied. In the case of oxazole phosphine oxides 8, the best results were obtained when the reaction was performed with chlorotrimethylsilane (1 M) and phenol (3 M) in dichloromethane²⁶ to give (2-aminomethyl-5-methyloxazol-4-yl) phosphine oxide 11aa $(R^1 = CH_3, R^2 = R^3 = H)$ and enantiomerically pure oxazole (-)-11ae derived from (S)-alanine ($R^1 = CH_3$, $R^2 = H$, $R^3 = CH_3$) in good yields (Scheme 3, Table 2, entries 10 and 11). However, in the case of oxazole phosphonate 10ad the deprotection was accomplished²⁷ with HCl 3 M in refluxing AcOEt and optically active oxazole (+)-12ad derived from (R)-alanine $(R^1 = CH_3, R^2 = CH_3, R^3 = H)$ was obtained (Scheme 3, Table 2, entry 12).



Scheme 3.

Table 2. Phosphorylated oxazoles 8, 10, 11 and 12

Entry	Compound	R	\mathbb{R}^1	R ²	R ³	Yield (%) ^a	$[\alpha]_{D}^{22,b}$
1	8aa	C ₆ H ₅	CH ₃	Н	Н	68	_
2	(±)-8ab	C_6H_5	CH ₃	Hc	CH ₃ ^c	70	_
3	(+)-8ad	C_6H_5	CH ₃	CH ₃	Н	72	+43.4
4	(-)-8ae	C_6H_5	CH ₃	Н	CH ₃	66	-43.4
5	(-)- 8ah	C_6H_5	CH ₃	Н	CH ₂ C ₆ H ₅	73	-11.0
6	(±)-10ab	OC_2H_5	CH ₃	Hc	CH ₃ ^c	45	_
7	(+)-10ad	OC_2H_5	CH ₃	CH ₃	Н	56	+39.7
8	(−) -10ae	OC_2H_5	CH_3	Н	CH ₃	50	-39.7
9	(+) -10bd	OC_2H_5	C_2H_5	CH ₃	Н	46	+34.5
10	11aa	C_6H_5	CH ₃	Н	Н	81	_
11	(−) -11ae	C_6H_5	CH_3	Н	CH ₃	74	-8.0
12	(+)-12ad	OC_2H_5	CH ₃	CH ₃	Н	46	+15.0

^a Yields refer to isolated compounds.

^b Degrees (for concentration, see Section 4).

^c Racemic.

2.3. Synthesis of phosphorylated oxazoles 16 and 17 with peptide residues

The ring opening of azirines can be extended to N-protected peptides.²⁰ Treatment of 3-methyl-2H-azirinyl phosphine oxide 1a ($R^1 = CH_3$) with both a optically active N-protected dipeptide N-Boc-(S)-glycine-phenylalanine 13a (Pep=Gly-Phe) and with a N-protected tripeptide N-Boc-(S)-alanine-glycine-glycine 13b (Pep = Ala-Gly-Gly) at low temperature (-80°C) in THF led to the formation of optically active *a*-ketamides containing a phosphine oxide group in the α -position (–)and 14aa $(R^1 = CH_3, Pep = Gly-Phe)$ (+)-14ab $(R^1 = CH_3, Pep = Ala-Gly-Gly)$, which were obtained as 1:1 diastereoisomeric mixtures²¹ in moderate yields (Scheme 4, Table 3, entries 1 and 2). Spectroscopic data were in agreement with the assigned structure of compounds 14 and the formation of adducts 14 could be explained, as before (Scheme 2), by formal addition of the carboxylic acid moiety of the N-protected peptide to the reactive carbon-nitrogen azirine double bond to give an unstable aziridine intermediate, followed by ring opening of zwitterionic oxazolone. The process can also be extended to azirine-phosphonates. Heating at 70°C (2 h) azirine **6b** ($R^1 = C_2H_5$) with N-Boc-(S)glycine-phenylalanine 13a (Pep=Gly-Phe) gave a 1:1 diastereoisomeric mixture²¹ of optically active α ketamide containing a phosphonate group in the α position (-)-15ba (Scheme 4, Table 3, entry 3).²² This formation of phosphapeptides 14 and 15 can be regarded as a peptide chain elongation which introduces an α -ketamine containing a phosphine oxide or a phosphonate group to the C-terminal end of the peptide. Ketamide (-)-14aa was then treated with triphenylphosphine and hexachloroethane in the presence of triethylamine and optically active oxazole phosphine oxide containing a peptide residue (-)-16aa was obtained (Scheme 4, Table 3, entry 4). In a similar manner oxazole phosphonate (-)-17ba was prepared from α -ketamide (-)-15ba derived from phosphonate

(Scheme 4, Table 3, entry 5). The formation of enantiomerically pure functionalized oxazoles 16 and 17 could also be explained through a similar mechanism to that reported for oxazoles derived from amino acids 8 and 10 (Scheme 3). This strategy describes the first preparation of optically active peptide-based oxazoles containing phosphinoyl 16 or phosphoryl substituents 17.

3. Conclusions

In conclusion, the first asymmetric synthesis of optically active oxazoles containing amino alkyl residues and with phosphine oxides 8 and 11 or phosphonate groups 10 and 12 has been described. The process involves a two-step procedure involving the ring opening of 2Hazirines derived from phosphine oxides 1 or phosphonates 6 with N-protected amino acids 2, followed by dehydration and ring closure of the first-formed α ketamides 3 and 7. This strategy can also be extended to N-protected peptides 13 to give the first family of phosphorylated optically active oxazoles 16 and 17 with peptide side chains. Functionalized oxazoles containing amino acid 8-12 and peptide residues 16 and 17 as well as phosphorylated α -ketamides phosphapeptides 3, 7, 14, and 15 are important synthons in organic synthesis and for the preparation of biologically active compounds with interest in medicinal chemistry.^{3,4,18,19}

4. Experimental

4.1. General methods

Analytical TLC was performed with Merck silica gel 60 F_{254} plates. Visualization was accomplished by UV light. Flash chromatography was carried out using Merck silica gel 60 (230–400 mesh ASTM). Melting points were determined with an Electrothermal IA9100



Scheme 4.

Table 3. α -Ketamides 14 and 15 and phosphorylated oxazoles 16 and 17

Entry	Compound	R	\mathbb{R}^1	Peptide	Yield (%) ^a	$[\alpha]_{D}^{22,b}$
1	(<i>—</i>)-14aa	C ₆ H ₅	CH ₃	(S)-Gly-Phe	55	-10.7
2	(+)- 14ab	C_6H_5	CH ₃	(S)-Ala-Gly-Gly	51	+3.3
3	(-)- 15ba	OC ₂ H ₅	C ₂ H ₅	(S)-Gly-Phe	48	-3.9
4	(-) -16aa	$C_6 H_5$	CH ₃	(S)-Gly-Phe	52	-6.6
5	(—)-17ba	OC_2H_5	C_2H_5	(S)-Gly-Phe	50	-3.6

^a Yields refer to isolated compounds.

^b Degrees (for concentration, see Section 4).

digital melting point apparatus and are uncorrected. ¹H (300 MHz), ¹³C (75 MHz) and ³¹P NMR (120 MHz) spectra were recorded on a Varian VXR 300 MHz spectrometer using CDCl₃ solutions with TMS as an internal reference for ¹H and ¹³C NMR spectra and phosphoric acid (85%) as external standard for ³¹P NMR spectra. Low-resolution mass spectra (MS) were obtained at 50-70 eV by electron impact (EIMS) on Hewlett-Packard 5971 and 5973 spectrometers, and obtained by APCI on a Hewlett-Packard 1100 spectrometer. Infrared spectra (IR) were recorded using a Nicolet IRFT Magna 550 spectrometer. $[\alpha]_{D}^{22}$ were taken on a Perkin-Elmer 341 polarimeter using a Na/ HaI lamp. Elemental analyses were performed in a LECO CHNS-932 apparatus. Azirines 1a¹¹ and 6a,b^{12a} were synthesized according to literature procedures.

4.2. General procedure for synthesis of 2-*tert*-butoxycarbonylamino-3-alkyl-*N*-(1-diphenylphosphinoyl-2-oxopropyl)alkylamide 3

To a -80° C solution of 3-alkyl-2*H*-azirin-2-yl diphenylphosphine oxide 1 (5 mmol) in THF (5 ml), a solution of *N*-protected amino acid 2 (15 mmol) in THF (5 ml) was added under a nitrogen atmosphere. Then, the mixture was allowed to warm to room temperature and stirred for 1–4 days. The solvent was evaporated in vacuo, and the subsequent residue was purified by flash column chromatography eluting with AcOEt.

4.2.1. 2-tert-Butoxycarbonylamino-*N*-(1-diphenylphosphinoyl-2-oxopropyl)acetamide (3aa). Yield 1.57 g (73%) obtained as a white solid from compound 1a (1.28 g, 5 mmol) and *N*-tert-butoxycarbonylglycine 2a (2.63 g, 15 mmol) as described in the general procedure: mp 182–183°C; ¹H NMR (300 MHz, CDCl₃): δ 1.40 (s, 9H), 2.18 (s, 3H), 3.62 (m, 2H), 4.98 (d, ³J_{HH}=7.5 Hz, 1H), 5.77 (dd, ²J_{PH}=9.0 Hz, ³J_{HH}=7.5 Hz, 1H), 7.39–7.95 (m, 11H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 28.2, 29.8, 44.0, 60.7 (d, ¹J_{PC}=65.0 Hz), 80.1, 128.4–132.8 (m), 155.7, 169.4 (d, ³J_{PC}=4.0 Hz), 200.6 ppm; ³¹P NMR (120 MHz, CDCl₃): δ 30.8 ppm; IR (KBr): 3423, 3211, 1724, 1699, 1668, 1228, 1182 cm⁻¹; MS (CI): *m*/*z* 431 (M⁺+1, 7). Anal. calcd for C₂₂H₂₇N₂O₅P: C, 61.39; H, 6.32; N, 6.51. Found C, 61.43; H, 6.31; N, 6.53.

(±)-2-tert-Butoxycarbonylamino-N-(1-diphenyl-4.2.2. phosphinoyl-2-oxopropyl)propanamide (3ab). Yield 1.44 g (65%) obtained as a white solid from compound 1a (1.28 g, 5 mmol) and (\pm) -N-tert-butoxycarbonylalanine **2b** (2.84 g, 15 mmol) as described in the general procedure: mp 146-147°C (hexane/AcOEt); ¹H NMR (300 MHz, CDCl₃): δ 0.88 (d, ${}^{3}J_{HH} = 6.9$ Hz, 6H), 1.38 (s, 18H), 2.09 and 2.15 (2s, 6H), 4.03 (m, 2H), 5.02 (2d, ${}^{3}J_{\rm HH} = 7.2$ Hz, 2H), 5.76, 5.80 (2dd, ${}^{2}J_{\rm PH} = 9.6$ Hz, ${}^{3}J_{\rm HH} = 7.2$ Hz, 2H), 7.41–7.92 (m, 22H) ppm; ${}^{13}C$ NMR (75 MHz, CDCl₃): δ 17.8, 18.0, 28.2, 29.7, 49.8, 50.6, 60.4 (d, ${}^{1}J_{PC} = 66.0$ Hz), 80.0 and 80.2, 128.4–132.8 (m), 155.1, 172.7 (d, ${}^{3}J_{PC} = 4.5$ Hz), 200.3, 200.8 ppm; ${}^{31}P$ NMR (120 MHz, CDCl₃): δ 32.4 and 33.1 ppm; IR (KBr): 3423, 3310, 3211, 3070, 2979, 1725, 1715, 1672, 1202, 1175 cm⁻¹; MS (CI): m/z 445 (M⁺+1, 12). Anal. calcd for $C_{23}H_{29}N_2O_5P$: C, 62.15; H, 6.58; N, 6.30. Found C, 62.11; H, 6.60; N, 6.31.

4.2.3. (±)-2-tert-Butoxycarbonylamino-3-hydroxy-N-(1diphenylphosphinoyl-2-oxopropyl)propanamide (3ac). Yield 1.50 g (65%) obtained as a white solid from compound 1a (1.28 g, 5 mmol) and (±)-N-tert-butoxycarbonylserine 2c (3.08 g, 15 mmol) as described in the general procedure: mp 74–75°C (hexane/AcOEt); ¹H NMR (300 MHz, CDCl₃): δ 1.35 (s, 18H), 2.20 and 2.16 (2s, 6H), 3.40-3.49 and 3.69-3.77 (2m, 4H), 4.08 (t, ${}^{3}J_{\rm HH} = 7.5$ Hz, 2H), 5.36 and 5.53 (2d, ${}^{3}J_{\rm HH} = 6.6$ Hz, 2H), 5.80 (2dd, ${}^{2}J_{PH}$ =9.6 Hz, ${}^{3}J_{HH}$ =6.6 Hz, 2H), 7.41– 7.90 (m, 22H) ppm; 13 C NMR (75 MHz, CDCl₃): δ 28.2, 29.5, 55.7 and 56.0, 60.6 (d, ${}^{1}J_{PC}$ =67.5 Hz), 62.4 and 62.7, 80.0, 127.8–132.8 (m), 155.6, 171.1 (d, ${}^{3}J_{PC} =$ 13.6 Hz), 200.8 ppm; ³¹P NMR (120 MHz, CDCl₃): δ 30.7 and 31.2 ppm; IR (KBr): 3525, 3429, 3310, 3197, 3051, 2979, 1724, 1712, 1664, 1175 cm⁻¹; MS (CI): m/z461 (M⁺+1, 13). Anal. calcd for C₂₃H₂₉N₂O₆P: C, 59.99; H, 6.35; N, 6.08. Found C, 59.92; H, 6.33; N, 6.09.

4.2.4. (+)-2-(*R*)-tert-Butoxycarbonylamino-*N*-(1-diphenyl-phosphinoyl-2-oxopropyl)propanamide (3ad). Yield 1.29 g (58%) obtained as a white solid from compound 1a (1.28 g, 5 mmol) and *N*-tert-butoxycarbonyl-(*R*)-alanine 2d (2.84 g, 15 mmol) as described in the general procedure: $[\alpha]_{D}^{22} = +34.0$ (*c* 1.00, CH₂Cl₂). For spectroscopic data see compound 3ab.

4.2.5. (-)-2-(*S*)-*tert*-Butoxycarbonylamino-*N*-(1-diphenyl-phosphinoyl-2-oxopropyl)propanamide (3ae). Yield 1.38 g (62%) obtained as a white solid from compound 1a (1.28 g, 5 mmol) and *N*-*tert*-butoxycarbonyl-(*S*)-alanine **2e** (2.84 g, 15 mmol) as described in the general procedure: $[\alpha]_{D}^{22} = -34.0$ (*c* 1.00, CH₂Cl₂). For spectroscopic data see compound **3ab**.

4.2.6. (+)-2-(*R*)-*tert*-Butoxycarbonylamino-3-hydroxy-*N*-(1-diphenylphosphinoyl-2-oxopropyl)propamide (3af). Yield 0.64 g (28%) obtained as a white solid from compound 1a (1.28 g, 5 mmol) and *N*-*tert*-butoxycarbonyl-(*R*)-serine 2f (3.08 g, 15 mmol) as described in the general procedure: $[\alpha]_D^{22} = +32.4$ (*c* 0.87, CH₂Cl₂). For spectroscopic data see compound 3ac.

4.2.7. (-)-2-(*S*)-*tert*-Butoxycarbonylamino-3-hydroxy-*N*-(1-diphenylphosphinoyl-2-oxopropyl)propanamide (3ag). Yield 1.52 g (66%) obtained as a white solid from compound 1a (1.28 g, 5 mmol) and *N*-*tert*-butoxycarbonyl-(*S*)-serine 2g (3.08 g, 15 mmol) as described in the general procedure: $[\alpha]_D^{22} = -32.4$ (*c* 0.21, CH₂Cl₂). For spectroscopic data see compound 3ac.

4.2.8. (-)-2-(*S*)-*tert*-Butoxycarbonylamino-*N*-(1-diphenyl-phosphinoyl-2-oxopropyl)phenyl propanamide (3ah). Yield 1.46 g (56%) obtained as a white solid from compound 1a (1.28 g, 5 mmol) and *N*-*tert*-butoxycarbonyl-(*S*)-phenylalanine 2h (3.98 g, 15 mmol) as described in the general procedure: mp 63–64°C (hexane/AcOEt); $[\alpha]_D^{22} = -11.4$ (*c* 1.00, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.25 (s, 18H), 2.02 and 2.16 (2s, 6H), 2.69 (m, 4H), 4.23 (m, 2H), 4.85 (d, ³J_{HH} = 7.8 Hz,

2H), 5.76 and 5.80 (2dd, ${}^{2}J_{\rm PH}$ =9.9 Hz, ${}^{3}J_{\rm HH}$ =7.8 Hz, 2H), 7.91–7.12 (m, 32H) ppm; 13 C NMR (75 MHz, CDCl₃): δ 28.1, 29.7, 37.8, 55.2 and 55.9, 60.1 (d, ${}^{1}J_{\rm PC}$ =66.5 Hz), 79.9, 126.6–136.3 (m), 155.1, 171.5 (d, ${}^{3}J_{\rm PC}$ =13.5 Hz), 200.9 ppm; 31 P NMR (120 MHz, CDCl₃): δ 31.6 and 32.5 ppm; IR (KBr): 3443, 3283, 3175, 1719, 1704, 1666, 1367, 1176 cm⁻¹; MS (CI): m/z521 (M⁺+1, 27). Anal. calcd for C₂₉H₃₃N₂O₅P: C, 66.91; H, 6.39; N, 5.38. Found C, 66.86; H, 6.39; N, 5.39.

4.3. General procedure for synthesis of diethyl 3-alkyl-(2-*tert*-butoxycarbonylaminoalkanoylamino)-2oxoalkylphosphonate 7

To diethyl 3-alkyl-2*H*-azirinyl phosphonate **6** (5 mmol) without solvent, the *N*-protected amino acid **2** (7.5 mmol) was added under a nitrogen atmosphere, at room temperature and with continuous stirring. The mixture was heated at 70°C for 2 h. Then, the subsequent residue was purified by flash column chromatography eluting with hexane/AcOEt affording **7** and a small proportion (10–20%) of pyrazine phosphonates.¹³

4.3.1. Diethyl (±)-1-(2-tert-butoxycarbonylaminopropanoylamino)-2-oxopropylphosphonate (7ab). Yield 1.29 g (68%) obtained as a colorless oil from compound 6a (0.96 g, 5 mmol) and (\pm) -N-tert-butoxycarbonylalanine **2b** (1.42 g, 7.5 mmol) as described in the general procedure: R_f 0.39 (AcOEt); ¹H NMR (300 MHz, CDCl₃): δ 1.28 (s, 12H), 1.39 (s, 18H), 2.36 (2s, 6H), 4.10 (m, 10H), 5.07 (2d, ${}^{3}J_{HH}$ =7.2 Hz, 2H), 5.76 (dd, ${}^{2}J_{PH}$ =23.3 Hz, ${}^{3}J_{HH}$ =8.5 Hz, 2H), 7.11 (d, ${}^{3}J_{HH}$ =8.5 Hz, 2H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 16.2, 18.3, 28.2, 28.9 (d), 49.9 (2s), 57.1 (2d, ${}^{1}J_{PC} = 140.5$ Hz), 63.6 (2s), 80.1, 155.2, 172.3 (d, ${}^{3}J_{PC} = 5.0$ Hz), 199.2 (d, $^{2}J_{PC} = 16.2$ Hz) ppm; ^{31}P NMR (120 MHz, CDCl₃): δ 15.7 and 15.8 ppm; IR (NaCl): 3270, 2861, 1725, 1709, 1679, 1590, 1165, 1113 cm⁻¹; MS (CI): m/z 381 (M⁺+1, 5). Anal. calcd for C₁₅H₂₉N₂O₇P: C, 47.36; H, 7.68; N, 7.36. Found C, 47.42; H, 7.67; N, 7.36.

4.3.2. Diethyl (+)-1-(2-(*R*)-*tert*-butoxycarbonylaminopropanoylamino)-2-oxopropylphosphonate (7ad). Yield 1.35 g (71%) obtained as a colorless oil from compound **6a** (0.96 g, 5 mmol) and *N*-*tert*-butoxycarbonyl-(*R*)alanine **2d** (1.42 g, 7.5 mmol) as described in the general procedure: $[\alpha]_{D}^{22} = +21.0$ (*c* 1.75, CH₂Cl₂). For spectroscopic data see compound **7ab**.

4.3.3. Diethyl (–)-1-(2-(*S***)-***tert***-butoxycarbonylaminopropanoylamino)-2-oxopropylphosphonate (7ae). Yield 1.25 g (66%) obtained as a colorless oil from compound 6a** (0.96 g, 5 mmol) and *N*-*tert*-butoxycarbonyl-(*S*)-alanine **2e** (1.42 g, 7.5 mmol) as described in the general procedure: $[\alpha]_{D}^{22} = -21.0$ (*c* 1.75, CH₂Cl₂). For spectroscopic data see compound **7ab**.

4.3.4. Diethyl (-)-1-(2-(S)-tert-butoxycarbonylamino-3hydroxypropanoylamino)-2-oxopropyl phosphonate (7ag). Yield 1.04 g (42%) obtained as a colorless oil from compound **6a** (0.96 g, 5 mmol) and *N*-tert-butoxycarbonyl-(S)-serine **2g** (1.54 g, 7.5 mmol) as described in the general procedure: R_f 0.33 (AcOEt/MeOH 10%); [α]²²_{pd} = -11.0 (*c* 0.39, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.26 (m, 12H), 1.39 (s, 18H), 2.32 (2s, 6H), 3.80–4.26 (m, 14H), 4.55 (s, 2H), 5.22 (dd, ²J_{PH}=23.3 Hz, ³J_{HH}=8.4 Hz, 2H), 5.71 (d, ³J_{HH}=7.5 Hz, 2H), 7.61 (d, ³J_{HH}=8.4 Hz, 2H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 16.2, 28.2, 28.7, 55.7 (2s), 58.2 (d, ¹J_{PC}=141.0 Hz), 62.9, 64.0, 80.4, 155.9, 171.2 (d, ³J_{PC}=5.0 Hz), 199.2 (d, ²J_{PC}=27.7 Hz) ppm; ³¹P NMR (120 MHz, CDCl₃): δ 16.2 and 16.6 ppm; IR (NaCl): 3310, 2924, 2852, 1723, 1710, 1699, 1591, 1164 cm⁻¹; MS (CI): *m*/*z* 397 (M⁺+1, 4). Anal. calcd for C₁₅H₂₉N₂O₈P: C, 45.45; H, 7.37; N, 7.07. Found C, 45.40; H, 7.38; N, 7.08.

4.3.5. Diethyl (+)-1-(2-(R)-tert-butoxycarbonylaminopropanoylamino)-2-oxobutylphosphonate (7bd). Yield 1.42 g (72%) obtained as a colorless oil from compound **6b** (1.03 g, 5 mmol) and *N*-tert-butoxycarbonyl-(R)alanine 2d (1.42 g, 7.5 mmol) as described in the general procedure: $R_{\rm f}$ 0.48 (AcOEt); $[\alpha]_{\rm D}^{22} = +24.4$ (c 0.47, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.03 (t, ${}^{3}J_{\rm HH} = 7.2$ Hz, 6H), 1.18–1.36 (m, 18H), 1.39 (s, 18H), 2.52 and 2.95 (2m, 4H), 4.12 (m, 10H), 4.98 (d, ${}^{3}J_{HH}$ 7.5 Hz, 2H), 5.18 (dd, ${}^{2}J_{PH}$ =22.7 Hz, ${}^{3}J_{HH}$ =8.4 Hz, 2H), 7.07 (d, ${}^{3}J_{HH}$ =8.5 Hz, 2H) ppm; 13 C NMR (75 MHz, CDCl₃): δ 7.5, 16.2, 18.2, 28.2, 34.9 (d, ${}^{3}J_{PC}$ = 3.5 Hz), 50.0 (2s), 57.0 (2d, ${}^{1}J_{PC}$ = 139.5 Hz), 63.5, 80.2, 155.3, 172.2 (d, ${}^{3}J_{PC}$ = 5.5 Hz), 202.4 (d, ${}^{2}J_{PC}$ = 13.6 Hz) ppm; ³¹P NMR (120 MHz, CDCl₃): δ 16.0 and 16.1 ppm; IR (NaCl): 3287, 2981, 2937, 2860, 1772, 1709, 1680, 1167, 977 cm⁻¹; MS (CI): m/z 395 (M⁺+1, 4). Anal. calcd for C₁₆H₃₁N₂O₇P: C, 48.72; H, 7.92; N, 7.10. Found C, 48.77; H, 7.93; N, 7.09.

4.3.6. Diethyl (-)-1-(2-(S)-tert-butoxycarbonylamino-3hydroxypropanoylamino)-2-oxobutylphosphonate (7bg). Yield 0.89 g (35%) obtained as a colorless oil from compound 6b (1.03 g, 5 mmol) and N-tert-butoxycarbonyl-(S)-serine 2g (1.54 g, 7.5 mmol) as described in the general procedure: $R_{\rm f}$ 0.50 (AcOEt); $[\alpha]_{\rm D}^{22} = -9.5$ (c 2.15, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.03 (t, ${}^{3}J_{\rm HH} = 7.2$ Hz, 6H), 1.27 (m, 12H), 1.40 (s, 18H), 2.54 and 2.83 (2m, 4H), 3.75-4.28 (m, 14H), 4.71 (s, 2H), 5.22 (dd, ${}^{2}J_{PH}$ =22.6 Hz, ${}^{3}J_{HH}$ =8.6 Hz, 2H), 5.72 (d, ${}^{3}J_{HH}$ =7.5 Hz, 2H), 7.60 (d, ${}^{3}J_{HH}$ =8.6 Hz, 2H) ppm; ${}^{13}C$ NMR (75 MHz, CDCl₃): δ 7.3, 16.2, 28.1, 34.7, 55.7 (2s), 57.1 (d, ${}^{1}J_{PC}$ =140.0 Hz), 62.8, 63.6, 80.2, 155.7, 170.9 (d, ${}^{3}J_{PC}$ =5.0 Hz), 202.1 (d, ${}^{2}J_{PC}$ =24.7 Hz) ppm; ³¹P NMR (120 MHz, CDCl₃): δ 16.4 and 16.8 ppm; IR (NaCl): 3250, 2938, 2860, 1725, 1711, 1679, 1382, 1165, 979 cm⁻¹; MS (CI): m/z 511 (M⁺+1, <1). Anal. calcd for C₁₆H₃₁N₂O₈P: C, 46.83; H, 7.61; N, 6.83. Found C, 46.78; H, 7.60; N, 6.84.

4.4. General procedure for synthesis of (5-alkyl-2-*tert*butoxycarbonylaminoalkyl)oxazol-4-yl diphenylphosphine oxide 8

To a room temperature solution of triphenylphosphine (1.70 g, 6.5 mmol) and hexachloroethane (1.54 g, 6.5 mmol), in THF a solution of 2-*tert*-butoxycarbonyl-amino-3-alkyl-*N*-(1-diphenylphosphinoyl-2-oxopropyl)-

alkylamide 3 was added under a nitrogen atmosphere. The mixture was stirred at that temperature for 5 min. After that, triethylamine (2.10 ml, 15 mmol) was added dropwise for 10 min. The subsequent mixture was heated at THF reflux for 20 h. The solvent was evaporated in vacuo and the residue was diluted with water and extracted with CH_2Cl_2 . The organic layers where dried over anhydrous $MgSO_4$, filtered and concentrated in vacuo. The crude residue was purified by flash column chromatography eluting with AcOEt.

4.4.1. (2-tert-Butoxycarbonylaminomethyl-5-methyl)oxazol-4-yl diphenylphosphine oxide (8aa). Yield 1.40 g (68%), obtained as a white solid from compound **3aa** (2.15 g, 5 mmol) as described in the general procedure: mp 104–105°C (hexane/AcOEt); ¹H NMR (300 MHz, CDCl₃): δ 1.39 (s, 9H), 2.52 (s, 3H), 4.34 (d, ³J_{HH}= 5.1 Hz, 2H), 5.28 (s, 1H), 7.82–7.36 (m, 10H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 11.6, 28.2, 37.8, 80.1, 126.3 (d, ¹J_{PC}=142.0 Hz), 128.3–133.2 (m), 155.5, 159.4 (d, ²J_{PC}=27.1 Hz), 160.3 (d, ³J_{PC}=17.0 Hz) ppm; ³¹P NMR (120 MHz, CDCl₃): δ 19.3 ppm; IR (KBr): 3264, 3078, 2980, 1712, 1553, 1275, 1188 cm⁻¹; MS (CI): *m*/*z* 413 (M⁺+1, 48). Anal. calcd for C₂₂H₂₅N₂O₄P: C, 64.07; H, 6.11; N, 6.79. Found C, 63.11; H, 6.12; N, 6.78.

4.4.2. (±)-[2-(1-tert-Butoxycarbonylaminoethyl)-5-methyl]oxazol-4-yl diphenylphosphine oxide (8ab). Yield 1.47 g (69%), obtained as a white solid from compound **3ab** (2.22 g, 5 mmol), as described in the general procedure: mp 120–121°C (hexane/AcOEt); ¹H NMR (300 MHz, CDCl₃): δ 1.37 (s, 9H), 1.43 (d, ³J_{HH}=7.2 Hz, 3H), 2.50 (s, 3H), 4.85 (q, ³J_{HH}=7.2 Hz, 1H), 5.00 (s, 1H), 7.48–7.34 (m, 10H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 11.7, 20.1, 28.3, 44.7, 80.0, 126.3 (d, ¹J_{PC}= 141.5 Hz), 128.3–133.6 (m), 154.9, 159.1 (d, ²J_{PC}=26.7 Hz), 163.8 (d, ³J_{PC}=17.1 Hz) ppm; ³¹P NMR (120 MHz, CDCl₃): δ 18.3 ppm; IR (KBr): 3337, 2992, 2945, 1719, 1540, 1175 cm⁻¹; MS (CI): *m*/*z* 427 (M⁺+ 1, 33). Anal. calcd for C₂₃H₂₇N₂O₄P: C, 64.78; H, 6.38; N, 6.57. Found C, 64.71; H, 6.37; N, 6.58.

4.4.3. (+)-[2-(1-(*R*)-tert-Butoxycarbonylaminoethyl)-5methyl]oxazol-4-yl diphenylphosphine oxide (8ad). Yield 1.53 g (72%), obtained as a white solid from compound **3ad** (2.22 g, 5 mmol), as described in the general procedure: $[\alpha]_{D}^{22} = +43.4$ (*c* 1.00, CH₂Cl₂). For spectroscopic data see compound **8ab**.

4.4.4. (-)-[2-(1-(*S*)-tert-Butoxycarbonylaminoethyl)-5methyl]oxazol-4-yl diphenylphosphine oxide (8ae). Yield 1.41 g (66%), obtained as a white solid from compound **3ae** (2.22 g, 5 mmol), as described in the general procedure: $[\alpha]_{D}^{22} = -43.4$ (*c* 1.00, CH₂Cl₂). For spectroscopic data see compound **8ab**.

4.4.5. (-)-[2-(1-(*S*)-*tert*-Butoxycarbonylamino-2-phenylethyl)-5-methyl]oxazol-4-yl diphenylphosphine oxide (8ah). Yield 1.83 g (73%), obtained as a white solid from compound 3ah (2.60 g, 5 mmol), as described in the general procedure: mp 115–116°C (hexane/AcOEt); [α]_D²² = -11.0 (*c* 1.00, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.34 (s, 9H), 2.51 (s, 3H), 3.11 (d, ³J_{HH} = 7.2 Hz, 2H), 5.09 (q, ³J_{HH} = 7.2 Hz, 1H), 5.16 (s, 1H), 6.90–7.73 (m, 15H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 11.5, 28.1, 39.8, 49.8, 79.9, 126.1 (d, ¹J_{PC} = 141.9 Hz), 126.7–135.7 (m), 154.8, 159.2 (d, ²J_{PC} = 26.7 Hz), 162.0 (d, ³J_{PC} = 17.1 Hz) ppm; ³¹P NMR (120 MHz, CDCl₃): δ 18.8 ppm; IR (KBr): 3250, 3058, 2979, 2939, 1719, 1188 cm⁻¹; MS (CI): m/z 503 (M⁺+1, 45). Anal. calcd for C₂₉H₃₁N₂O₄P: C, 69.31; H, 6.22; N, 5.57. Found C, 69.39; H, 6.21; N, 5.56.

4.5. General procedure for synthesis of diethyl [5-alkyl-2-(1-tert-butoxycarbonylaminoalkyl)]oxazol-4-yl phos-phonate 10

To a room temperature solution of triphenylphosphine (1.70 g, 6.5 mmol) and hexachloroethane (1.54 g, 6.5 mmol), in toluene a solution of diethyl 3-alkyl-(2-*tert*-butoxycarbonylaminoalkanoylamino)-2-oxoalkylphosphonate **7** was added under a nitrogen atmosphere. The mixture was stirred at that temperature for 5 min. After that, triethylamine (2.10 ml, 15 mmol) was added dropwise slowly for 10 min. The subsequent mixture was heated at toluene reflux and stirred for 20 h. The solvent was evaporated in vacuo and the residue was purified by precipitating in cold ethyl ether. The ethereal layers were concentrated in vacuo and the residue was ground with cold water and filtered. The aqueous layer was concentrated again affording the compounds **10** as colorless oils.

4.5.1. Diethyl (±)-[2-(1-*tert*-butoxycarbonylaminoethyl)-**5-methyl]oxazol-4-yl phosphonate (10ab)**. Yield 0.81 g (45%), obtained as a colorless oil from compound **7ab** (1.90 g, 5 mmol), as described in the general procedure method: $R_{\rm f}$ 0.45 (AcOEt); ¹H NMR (300 MHz, CDCl₃): δ 1.29 (t, ³J_{HH}=7.0 Hz, 6H), 1.38 (s, 9H), 1.45 (d, ³J_{HH}=7.0 Hz, 3H), 2.49 (d, ⁴J_{HH}=1.8 Hz, 3H), 4.08 (m, 4H), 4.84 (q, ³J_{HH}=7.0 Hz, 1H), 5.14 (s, 1H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 11.4, 16.2 (d, ³J_{PC}=6.5 Hz), 20.2, 28.2, 44.7, 62.5 (d, ²J_{PC}=5.5 Hz), 79.9, 124.0 (d, ¹J_{PC}=41.5 Hz), 154.8, 158.5 (d, ²J_{PC}=39.3 Hz), 164.1 (d, ³J_{PC}=21.2 Hz) ppm; ³¹P NMR (120 MHz, CDCl₃): δ 9.6 ppm; IR (NaCl): 3260, 2917, 2861, 1700, 1521, 1162, 1027 cm⁻¹; MS (CI): m/z 363 (M⁺+1, 7). Anal. calcd for C₁₅H₂₇N₂O₆P: C, 49.72; H, 7.51; N, 7.73. Found C, 49.79; H, 7.52; N, 7.72.

4.5.2. Diethyl (+)-[2-(1-(*R*)-*tert*-butoxycarbonylaminoethyl)-5-methyl]oxazol-4-yl phosphonate (10ad). Yield 1.01 g (56%), obtained as a colorless oil from compound 7ad (1.90 g, 5 mmol), as described in the general procedure: $R_{\rm f}$ 0.45 (AcOEt); $[\alpha]_{\rm D}^{22} = +39.7$ (*c* 0.3, CH₂Cl₂). For spectroscopic data see compound 10ab.

4.5.3. Diethyl (-)-[2-(1-(S)-tert-butoxycarbonylaminoethyl)-5-methyl]oxazol-4-yl phosphonate (10ae). Yield 0.91 g (50%), obtained as a colorless oil from compound 7ae (1.90 g, 5 mmol), as described in the general procedure: $R_{\rm f}$ 0.45 (AcOEt); $[\alpha]_{\rm D}^{22} = -39.7$ (c 0.3, CH₂Cl₂). For spectroscopic data see compound 10ab. 4.5.4. Diethyl (+)-[2-(1-(*R*)-tert-butoxycarbonylaminoethyl)-5-ethyl]oxazol-4-yl phosphonate (10bd). Yield 0.87 g (46%), obtained as a colorless oil from compound 7bd (1.97 g, 5 mmol), as described in the general procedure: $R_{\rm f}$ 0.40 (AcOEt); $[\alpha]_{\rm D}^{22}$ =+34.5 (*c* 0.33, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.19 (t, ³J_{HH}=7.5 Hz, 3H), 1.29 (t, ³J_{HH}=7.0 Hz, 6H), 1.38 (s, 9H), 1.45 (d, ³J_{HH}=7.0 Hz, 3H), 2.92 (dq, ³J_{HH}= 7.5 Hz, ⁴J_{HH}=1.8 Hz, 2H), 4.08 (m, 4H), 4.86 (q, ³J_{HH}=7.0 Hz, 1H), 5.17 (s, 1H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 12.6, 16.2 (d, ³J_{PC}=6.5 Hz), 19.1, 20.2, 28.2, 44.7, 62.5 (d, ²J_{PC}=5.5 Hz), 79.9, 123.3 (d, ¹J_{PC}=243.9 Hz), 154.8, 163.4 (d, ²J_{PC}=39.8 Hz), 164.1 (d, ³J_{PC}=21.2 Hz) ppm; ³¹P NMR (120 MHz, CDCl₃): δ 9.7 ppm; IR (NaCl): 3254, 2917, 2861, 1702, 1519, 1165, 1119 cm⁻¹; MS (CI): *m*/z 377 (M⁺+1, 10). Anal. calcd for C₁₆H₂₉N₂O₆P: C, 51.06; H, 7.77; N, 7.44. Found C, 50.94; H, 7.76; N, 7.45.

4.6. General procedure for synthesis of (5-alkyl-2aminoalkyl)oxazol-4-yl diphenylphosphine oxide 11

To a solution previously prepared and kept under anhydrous Na_2CO_3 of chlorotrimethylsilane 1 M and phenol 3 M in CH_2Cl_2 (15 mmol) a solution of protected oxazole 8 was added at room temperature and under a nitrogen atmosphere. The mixture was stirred at that temperature for 24 h. Then, it was washed with a NaOH 2N solution (10 ml) and extracted with CH_2Cl_2 . The organic layer was dried over anhydrous MgSO₄, filtered and the solvent was evaporated under vacuum. The crude residue was ground in hexane/ ether and, after filtering, was washed with the same mixture of solvents. The products were crystallized from hexane/AcOEt.

4.6.1. (2-Aminomethyl-5-methyl)oxazol-4-yl diphenylphosphine oxide (11aa). Yield 1.26 g (81%), obtained as a white solid from compound **8aa** (2.06 g, 5 mmol) as described in the general procedure: mp 131–132°C (hexane/AcOEt); ¹H NMR (300 MHz, CDCl₃): δ 1.66 (s, 2H), 2.57 (d, ⁴J_{PH}=1.8 Hz, 3H), 3.89 (s, 2H), 7.39–7.85 (m, 10H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 11.7, 39.4, 126.2 (d, ¹J_{PC}=142.0 Hz), 128.3–133.6 (m), 159.1 (d, ²J_{PC}=26.7 Hz), 163.9 (d, ³J_{PC}=16.6 Hz) ppm; ³¹P NMR (120 MHz, CDCl₃): δ 18.7 ppm; IR (KBr): 3376, 3303, 2932, 1593, 1440, 1195, 1129 cm⁻¹; MS (EI): m/z 312 (M⁺, 77). Anal. calcd for C₁₇H₁₇N₂O₂P: C, 65.38; H, 5.49; N, 8.97. Found C, 65.31; H, 5.49; N, 8.98.

4.6.2. (-)-[2-(1-(*S*)-Aminoethyl)-5-methyl]oxazol-4-yl diphenylphosphine oxide (11ae). Yield 1.21 g (74%), obtained as a white solid from compound **8ae** (2.22 g, 5 mmol), as described in the general procedure: mp 126–127°C (hexane/AcOEt); $[\alpha]_{D}^{22} = -8.0$ (*c* 0.3, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.46 (d, ${}^{3}J_{HH} = 6.9$ Hz, 3H), 1.87 (s, 2H), 2.51 (s, 3H), 4.07 (q, ${}^{3}J_{HH} = 6.9$ Hz, 1H), 7.36–7.82 (m, 10H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 11.7, 21.6, 45.5, 125.9 (d, ${}^{1}J_{PC} = 142.1$ Hz), 128.3–133.6 (m), 159.0 (d, ${}^{2}J_{PC} = 26.7$ Hz), 166.9 (d, ${}^{3}J_{PC} = 17.1$ Hz) ppm; ³¹P NMR (120 MHz, CDCl₃): δ 18.8 ppm; IR (KBr): 3350, 3283, 3058, 1593, 1188, 1173 cm⁻¹; MS (EI): m/z 326 (M⁺,

4.7. Synthesis of diethyl (+)-2-(*R*)-aminoethyl)-5-methyloxazol-4-yl phosphonate (12ad)

To a solution of compound 10ad (1.81 g, 5 mmol) in AcOEt, a solution of HCl 3 M (5 ml, 15 mmol) was added dropwise slowly. The mixture was heated at AcOEt reflux for 15 h. Then, the solution was extracted with water and the aqueous layer was concentrated in vacuo. The crude residue was purified by silica gel column chromatography eluting with AcOEt/ MeOH affording compound 12ad. 0.60 g (46%), obtained as a white solid from compound **11ad** (1.81) g, 5 mmol) as described in the general procedure: $R_{\rm f}$ 0.25 (AcOEt/MeOH 25%); $[\alpha]_{D}^{22} = +15.0$ (c 0.10, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.35 (t, ³J_{HH} = 6.8 Hz, 6H), 1.77 (dd, ³J_{HH} = 6.1 Hz, ³J_{NH} = 42.4 Hz, 3H), 1.78 (s, 2H), 2.51 (s, 3H), 4.20 (m, 5H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 11.5, 16.2 (d, ³J_{PC}=6.5 Hz), 17.5, 45.1, 63.2 (d, ²J_{PC}=5.0 Hz), 127.6 (d, ¹J_{PC}=226.1 Hz), 158.4 (d, ²J_{PC}=36.3 Hz), 160.0 (d, ${}^{3}J_{PC} = 22.1$ Hz) ppm; ${}^{31}P$ NMR (120 MHz, CDCl₃): δ 10.6 ppm; IR (KBr): 3383, 3269, 1646, 1228, 1036 cm⁻¹; MS (CI): m/z 263 (M⁺+1, 82). Anal. calcd for C₁₀H₁₉N₂O₄P: C, 45.80; H, 7.30; N, 10.68. Found C, 45.90; H, 7.31; N, 10.66.

4.8. General procedure for synthesis of *N*-Boc-peptides 13 (see Ref. 3)

4.8.1. (+)-2-(*S*)-[*N*-(2-*tert*-Butoxycarbonylamino)acetylamino]-3-phenylpropionic acid 13a. Yield 1.29 g (80%), obtained as a white solid from Gly-Phe (1.11 g, 5 mmol) as described in the general procedure: mp 141– 142°C (hexane/diethyl ether); $[\alpha]_{D}^{22}$ =+31.2 (*c* 1.00, MeOH); ¹H NMR (300 MHz, CDCl₃): δ 1.36 (s, 9H), 2.97 (q, ³J_{HH}=5.4 Hz, ²J_{HHgem}=13.5 Hz, 1H), 3.09 (q, ³J_{HH}=5.4 Hz, ²J_{HHgem}=13.5 Hz, 1H), 3.59 (q, ³J_{HH}= 6.3 Hz, ²J_{HHgem}=16.8 Hz, 1H), 3.79 (q, ³J_{HH}=6.3 Hz, ²J_{HHgem}=16.8 Hz, 1H), 4.77 (t, ³J_{HH}=5.4 Hz, 1H), 5.42 (s, 1H), 6.81 (d, ³J_{HH}=6.3 Hz, 1H), 7.07–7.20 (m, 5H), 9.21 (s, 1H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 28.2, 37.3, 43.6, 45.1, 80.5, 127.0–135.8 (m), 156.3, 170.0 and 173.5 ppm; IR (KBr): 3469, 3370, 3032, 1728, 1626. 1560 cm⁻¹; MS (CI): *m*/*z* 341 (M⁺+3, 2), 339 (M⁺+1, 7). Anal. calcd for C₁₇H₂₆N₂O₅: C, 60.34; H, 7.74; N, 8.28. Found: C, 60.24; H, 7.79; N, 8.29.

4.8.2. (-)-2-(*S*)-[*N*-(2-*tert*-Butoxycarbonylaminopropionylamino)acetylamino]acetic acid 13b. Yield 0.97 g (64%), obtained a a white solid from Ala-Gly-Gly (1.02 g, 5 mmol) as described in the general procedure: mp 76–77°C (hexane/diethyl ether); $[\alpha]_D^{22} = -7.9$ (*c* 0.52, MeOH); ¹H NMR (300 MHz, CDCl₃): δ 1.18 (d, ${}^{3}J_{\rm HH} = 7.2$ Hz, 3H), 1.36 (s, 9H), 3.67 (s, 2H), 3.74 (d, ${}^{2}J_{\rm HHgem} = 3.0$ Hz, 2H), 3.88 (q, ${}^{3}J_{\rm HH} = 7.2$ Hz, 1H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 17.9, 28.7, 43.5, 52.2, 66.9, 80.9, 158.1, 171.7, 176.5 ppm; IR (KBr): 3400, 3356, 3310, 1750, 1692, 1672, 1626, 1527 cm⁻¹; MS (CI): *m/z* 304 (M⁺+1, 8). Anal. calcd for C₁₂H₂₁N₃O₆: C, 47.52; H, 6.98; N, 13.85. Found C, 47.47; H, 6.99; N, 13.87.

4.9. General procedure for synthesis of peptide containing α -ketamides 14

To a solution of 3-alkyl-2*H*-azirinyl-2-phosphine oxide 1 (1.28 g, 5 mmol) in THF (5 ml), a solution of *N*-Boc-peptide 13 (15 mmol) in THF (5 ml) is added under a nitrogen atmosphere, at -80° C and with continuous stirring. The mixture was stirred allowing to warm to room temperature for 4 days. The solvent was concentrated under vacuum, and the residue was purified by flash column chromatography eluting with AcOEt.

(-)-2-(S)-(2-tert-Butoxycarbonylaminoethanoyl-4.9.1. amino) - N - (1 - diphenylphosphinoyl - 2 - oxopropyl) - 3phenylpropanamide (14aa). Yield 1.18 g (41%) obtained as a white solid from N-Boc-(S)-GlyPhe 13a (4.83 g, 15 mmol) as described in the general procedure: mp:110-111°C; $[\alpha]_D^{22} = -10.7$ (c 0.68, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.39 (s, 18H), 1.99 and 2.11, (2s, 6H), 2.58 (d, ${}^{3}J_{HH}$ =6.3 Hz, 4H), 2.74 (dd, ${}^{3}J_{HH}$ =7.2 Hz, ${}^{2}J_{HHgem}$ =13.8 Hz, 2H), 2.87 (dd, ${}^{3}J_{HH}$ =6.9 Hz, ${}^{2}J_{HHgem}$ =13.8 Hz, 2H), 3.63 (m, 4H), 4.70 and 4.80 (dq, ${}^{3}J_{HH}$ =7.2 Hz, 2H), 5.34 and 5.41 (2s, 2H), 5.84 and 5.76 (2dd, ${}^{2}J_{PH}$ =11.4 Hz, ${}^{3}J_{HH}$ =9.3 Hz, 2H), 6.45 and 6.75 (2d, ${}^{3}J_{HH}$ =6.9 Hz, 2H), 6.87–8.06 (m, 32H) prm: ${}^{13}C$ NMP (75 MHz, CDC1); δ 28.2, 20.7, 37.0 ppm; ¹³C NMR (75 MHz, CDCl₃): δ 28.3, 29.7, 37.9, 44.0, 54.0 (2s), 60.4 (2d, ${}^{1}J_{PC} = 65.5$ Hz), 80.0 (2s), 126.8–136.1 (m), 155.9, 169.4 and 170.9 (2d, ${}^{3}J_{PC} = 4.0$ Hz), 200.6 (2s) ppm; ³¹P NMR (120 MHz, $CDCl_3$): δ 31.2 and 31.7 ppm; IR (KBr): 3277, 3071, 2979, 2939, 1732, 1724, 1708, 1659, 1540, 1169 cm⁻¹; MS (CI): m/z578 (M⁺+1, 26). Anal. calcd for C₃₁H₃₆N₃O₆P: C, 64.46; H, 6.28; N, 7.27. Found C, 64.44; H, 6.29; N, 7.28.

4.9.2. (+)-2-[-2-(S)-(2-tert-Butoxycarbonylamino)propanoylamino]ethanoylamino - N - (1 - diphenylphosphinoyl-2-oxopropyl)acetamide (14ab). Yield 1.06 g (38%) obtained as a white solid from N-Boc-(S)-AlaGlyGly (4.55 g, 15 mmol) as described in the general procedure: mp 66–67°C (hexane/AcOEt); $[\alpha]_D^{22} = +3.3$ (c 1.63, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.33 (m, 24H), 2.02 (s, 6H), 2.53 (s, 2H), 3.59-4.06 (m, 8H), 4.20 (dq, ${}^{3}J_{HH} = 6.2$ Hz, ${}^{3}J_{HH} = 6.7$ Hz, 2H), 5.61 (d, ${}^{3}J_{HH} = 6.0$ Hz, 2H), 5.76 (dd, ${}^{2}J_{PH} = 10.3$ Hz, ${}^{3}J_{HH} = 9.4$ Hz, 2H), 7.28 (s, 2H), 7.38–7.87 (m, 20H), 8.13 (s, 2H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 18.5, 28.3, 29.7, 42.9, 50.5, 60.8 (d, ${}^{1}J_{PC} = 64.5$ Hz), 80.0, 128.4–132.9 (m), 155.8, 168.8, 169.6 (d, ${}^{3}J_{PC} = 4.0$ Hz), 173.9, 200.8 ppm; ³¹P NMR (120 MHz, $CDCl_3$): δ 31.4 and 31.5 ppm; IR (KBr): 3265, 3071, 2975, 2928, 1731, 1724, 1708, 1657, 1540, 1175 cm⁻¹; MS (CI): m/z 559 (M⁺+1, 5). Anal. calcd for C₂₇H₃₅N₄O₇P: C, 58.06; H, 6.32; N, 10.03. Found C, 57.93; H, 6.31; N, 10.03.

4.10. Synthesis of diethyl (-)-1-[2-(S)-(2-*tert*-butoxycarbonylaminoethanoylamino)-3-phenylpropanoylamino]-2-oxobutylphosphonate (15ba)

To diethyl 3-alkyl-2H-azirinyl phosphonate 6 (5 mmol) without solvent, the *N*-Boc-peptide 13 (7.5 mmol) is added under a nitrogen atmosphere, at room temperature and with continuous stirring. The mixture was

heated at 70°C for 2 h and the subsequent residue was purified by flash column chromatography eluting with AcOEt affording a small proportion (10–20%) of pyrazine phosphonates¹³ and **15ba** (1.13 g, 43%) obtained as colorless oil from N-Boc-(S)-GlyPhe 13a (4.83 g, 15 mmol), as described in the general procedure: $R_{\rm f}$ 0.37; $[\alpha]_{D}^{22} = -3.9$ (c 1.00, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 0.97 and 0.98 (2t, ${}^{3}J_{HH}$ = 7.0 Hz, 6H), 1.21 (m, 12H), 1.37 and 1.38 (2s, 18H), 2.38–3.12 (m, 8H), 3.71 (m, 4H), 3.96-4.25 (m, 10H), 5.18-5.11 (2dd, ${}^{2}J_{PH}$ = 22.0 and 21.5 Hz, ${}^{3}J_{HH}$ = 8.2 and 8.5 Hz, 2H), 7.12–7.91 (m, 14H) ppm; ${}^{13}C$ NMR (75 MHz, CDCl₃): δ 7.4, 16.2, 28.2, 34.7 (d, ${}^{3}J_{PC} = 15.6$ Hz), 37.9 and 38.2, 44.1, 53.8 and 54.1 (2s), 57.1 (d, ${}^{1}J_{PC} = 141.5$ Hz), 63.5, 80.0 (2s), 126.8-136.2 (m), 155.9, 169.5 and 169.7 (2s), 170.4 and 170.5 (2d, ${}^{3}J_{PC}$ =5.0 and 6.0 Hz), 202.4 (d, ${}^{2}J_{PC}$ =26.7 Hz) ppm; ${}^{31}P$ NMR (120 MHz, CDCl₃): δ 16.0 and 16.1 ppm; IR (NaCl): 3255, 3065, 2983, 2935, 1730, 1724, 1708, 1682, 1165 cm⁻¹; MS (CI): m/z 528 $(M^++1, 3)$. Anal. calcd for $C_{24}H_{38}N_3O_8P$: C, 54.64; H, 7.26; N, 7.97. Found C, 54.69; H, 7.25; N, 7.96.

4.11. General procedures for synthesis of peptide containing oxazoles 16 and 17

To a room temperature solution of triphenylphosphine (1.70 g, 6.5 mmol) and hexachloroethane (1.54 g, 6.5 mmol), in toluene a solution of peptide containing α -ketamides 14 and 15 was added under a nitrogen atmosphere. The mixture was stirred at that temperature for 5 min. After that, triethylamine (2.10 ml, 15 mmol) was added dropwise for 10 min. The mixture was heated at toluene reflux for 1–3 days. The solvent was evaporated in vacuo and the residue was diluted with water and extracted with CH₂Cl₂. The organic layers where dried over anhydrous magnesium sulfate, filtered and concentrated in vacuo. The crude residue was purified by flash column chromatography eluting with AcOEt.

4.11.1. (-)-2-[(1-(*S*)-*tert*-Butoxycarbonylaminoethanoylamino)-2-phenylethyl]-5-methyloxazol-4-yl diphenylphosphine oxide 16aa. Yield 1.45 g (52%) obtained as a white solid from compound 14aa (2.89 g, 5 mmol) as described in the general procedure: mp 74–75°C; $[\alpha]_D^{22} =$ -6.6 (*c* 0.50, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.38 (s, 9H), 2.51 (s, 3H), 3.20 (t, ³J_{HH} = 5.8, 4.9 Hz, 2H), 3.75 (m, 2H), 5.18 (s, 1H), 5.47 (q, ³J_{HH} = 6.0, 7.2 Hz), 6.83–7.77 (m, 16H) ppm; ¹³C NMR (75 MHz, CDCl₃): δ 11.6, 28.2, 39.4, 44.3, 48.4, 80.2, 126.9–135.5 (m), 155.9, 159.3 (d, ²J_{PC} = 26.7 Hz), 161.5 (d, ³J_{PC} = 17.1 Hz), 169.0 ppm; ³¹P NMR (120 MHz, CDCl₃): δ 19.6 ppm; IR (KBr): 3277, 3071, 2979, 2939, 1732, 1724, 1708, 1659, 1540, 1169 cm⁻¹; MS (CI): *m*/*z* 560 (M⁺+1, 53). Anal. calcd for C₃₁H₃₄N₃O₅P: C, 66.55; H, 6.12; N, 7.51. Found C, 66.48; H, 6.11; N, 7.51.

4.11.2. Diethyl (-)-2-[(1-(*S*)-*tert*-butoxycarbonylaminoethanoylamino)-2-phenylethyl]-5-ethyloxazol-4-yl phosphonate 17ba. Yield 1.27 g (50%) obtained as a colorless oil from compound 15ba (2.63 g, 5 mmol) as described in the general procedure: $R_{\rm f}$ 0.38; $[\alpha]_{\rm D}^2 = -3.6$ (*c* 0.30, CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃): δ 1.19 (t, ³J_{HH}= 7.5 Hz, 3H), 1.30 (q, ${}^{3}J_{\rm HH}$ =7.1 Hz, 6H), 1.45 (s, 9H), 2.93 (m, 2H), 3.20 (d, ${}^{3}J_{\rm HH}$ =6.3 Hz, 2H), 3.82 (m, 2H), 4.08 (m, 4H), 5.12 (s, 1H), 5.48 (q, ${}^{2}J_{\rm HH}$ =7.0, 1H), 6.74–7.26 (m, 6H) ppm; 13 C NMR (75 MHz, CDCl₃): δ 12.7, 16.2 (d, ${}^{3}J_{\rm PC}$ =4.5 Hz), 19.1, 28.3, 40.0, 48.5, 62.5 (d, ${}^{2}J_{\rm PC}$ =4.5 Hz), 80.3, 123.0 (d, ${}^{1}J_{\rm PC}$ =241.2 Hz), 127.0–135 (m), 155.8, 161.8 (d, ${}^{3}J_{\rm PC}$ =20.7 Hz), 163.8 (d, ${}^{2}J_{\rm PC}$ =39.8 Hz), 168.9 ppm; 31 P NMR (120 MHz, CDCl₃): δ 10.2 ppm; IR (NaCl): 3283, 2979, 2925, 1712, 1672, 1513, 1367, 1255, 1162, 1029 cm⁻¹; MS (CI): m/z 510 (M⁺+1, 100). Anal. calcd for C₂₄H₃₆N₃O₇P: C, 56.57; H, 7.12; N, 8.25. Found C, 56.62; H, 7.12; N, 8.24.

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